

Review

Human papillomavirus, cervical carcinogenesis and chemoprevention with Indole derivatives – a review of pathomechanisms

Gudrun C. Rieck and Alison N. Fiander

Department of Obstetrics and Gynaecology, Wales College of Medicine, Cardiff University, Heath Park, Cardiff, UK

Cervical cancer is the second most common female cancer worldwide with high risk Human Papillomavirus (HPV) infection playing an essential aetiological role. Oestrogen interacts with HPV at a cellular level causing cell growth and inhibition of apoptosis. Indole derivatives, formed during digestion of cruciferous vegetables, have been shown to have chemopreventative properties inhibiting HPV transcription and influencing oestrogen metabolism. This review describes the interactions between HPV, oestrogen and indole derivatives. Further clinical research is required to evaluate the chemopreventative properties of these agents.

Keywords: Cervical cancer / Chemoprevention / HPV / Indole derivatives / Oestrogen

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1 Introduction

Cervical cancer is the second commonest female cancer worldwide with Human Papillomavirus (HPV) playing a central aetiological role [1, 2]. HPVs can be classified according to their carcinogenic potential; high-risk types being associated with cervical cancer and cervical intraepithelial neoplasia, whereas low-risk HPVs are rarely associated with anogenital neoplasia [3–8]. High-Risk (HR)-HPV DNA is found in 99.7% of cervical tumours [9]. A multi-centre study showed that infection with HR-HPV has an odds ratio (OR) of 158.2 (95%CI, 113.4–220.6) for cervical cancer [3]. HPV is also associated with other diseases including head and neck cancers [10, 11] and recurrent respiratory papillomatosis [11, 12]. Other factors such as smoking [14], oral contraceptive pill usage [15, 16], multiparity [17, 18] and chlamydial infection [19–22] are weak independent risk factors for cervical cancer.

Correspondence: Dr. Gudrun Christine Rieck, Obstetrics and Gynaecology, Cardiff University, Heath Park, Cardiff, CF14 4XN, UK
E-mail: g.rieck@doctors.net.uk
Fax: +44-0-2920707863

Abbreviations: CIN, cervical intraepithelial neoplasia; DIM, diindolylmethane; HR, high risk; HRT, hormone replacement therapy, HVP, human papillomavirus; IARC, international agency for research on cancer; I3C, indole-3-carbinol; OR, odds ratio; Rb, retinoblastoma

2 HPV and oestrogen in cervical carcinogenesis

Genital HPV infection is usually transmitted through sexual contact, although other HPV viruses can be transmitted by skin to skin contact, *e.g.* verruca-causing HPVs [23]. Infection with HPV may occur more easily in the cervical transformation zone where columnar epithelium undergoes squamous metaplasia, the epithelium being thinner and therefore more susceptible to HPV infection [24].

The transformation zone, the most oestrogen sensitive region of the cervix, is where the majority of cervical cancers develop [25]. Animal studies confirm the sensitivity of the cervical transformation zone to oestrogen-induced carcinogenesis [26, 27]. Expression of cervical oestrogen receptors is highest in the transformation zone, with receptor-positive cells localised mainly in the parabasal and intermediate cell layers [28]. It has been suggested that there is a difference in the oestrogen and/or progesterone receptor status between the normal cervix and the HPV infected cervix [29–33], although this was not confirmed in all studies [34]. Cells in cervical intraepithelial neoplasia (CIN) appear to lose their oestrogen expression [32, 35–37], whilst on the other hand, increased levels of 2-OH-oestrone may increase apoptosis in HPV-infected cells, thereby having a protective effect against the development of cervical cancer [38]. Women who express higher levels of oes-

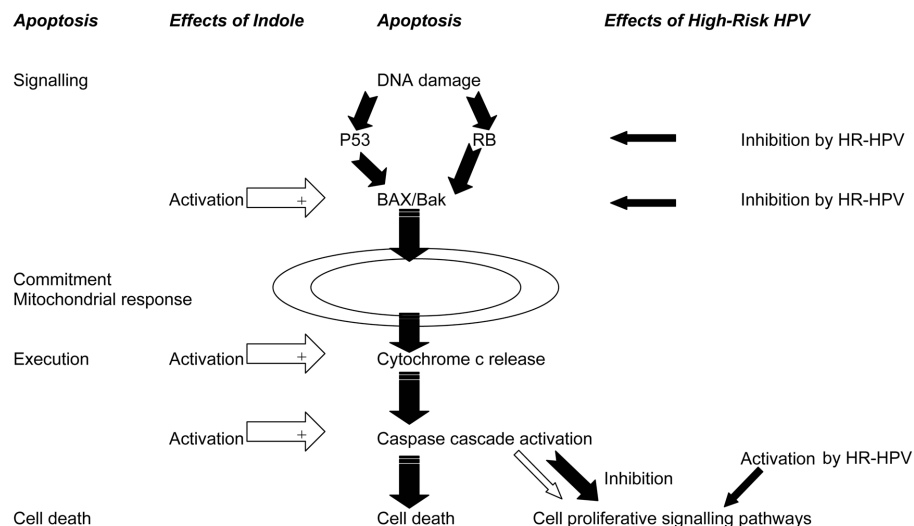


Figure 1. Simplified figure of apoptosis under influence of HR-HPV and chemopreventive agents such as Indoles. In general, DNA damage will activate tumour suppressor families such as p53 and Rb. They will prevent the cell from proceeding into S-phase. If the DNA damage cannot be repaired then pro-apoptotic factors such as Bax/Bak will be activated. This causes release of Cytochrome c from the mitochondria and the endoplasmic reticulum. Released cytochrome c activates the Caspase system, which finally executes cell death. High-Risk HPV (HR-HPV) inhibits tumour suppressors such as Rb thus releasing transcription factor E2F, which finally supports cell proliferation. HPV also inhibits apoptosis by activating anti-apoptotic factors and by inhibiting pro-apoptotic factors. Additionally, HPV activates cell signalling pathways, which cause cell proliferation. Indole derivatives on the other hand support apoptosis by stimulation of tumour suppressor genes such as BRCA, by activation of pro-apoptotic factors and by inhibition of proliferative cell signalling pathways.

trogen receptors are more likely to have cervical HPV infection suggesting that oestrogen receptors may play an important part in cervical HPV infection [31].

HPV related carcinogenesis occurs at several points in the cell cycle. HR-HPVs inhibit tumour suppressor families such as p53 and retinoblastoma protein (Rb) and inhibit cell death (apoptosis) at various steps in the apoptotic process (see Fig. 1).

The Rb tumour suppressor protein family function as negative regulators of the cell cycle. In its active state it binds to E2F transcription factors. Responding to cell growth-promoting factors, cyclin-dependent kinases inactivate the Rb-E2F complex through phosphorylation of the Rb protein releasing E2F, which induces progression through the cell cycle. The HPV oncoprotein E7 binds to the Rb-E2F complex, thereby inactivating the Rb tumour suppressor, releasing E2F transcription factors. This leads to stimulation of other genes such as p16 and p14. Both regulate an ubiquitin ligase which controls levels of the tumour suppressor p53 (see review by Doorbar 2006 [39]). The HR-HPV protein E6 associates with p53, mediating p53 ubiquitination and degradation [40]. E6 can also inhibit apoptosis by binding to Bak [41] and Bax [42, 43].

Other HPV proteins, such as E5, may contribute to the carcinogenesis of cervical cancer but the exact role is poorly understood. E5 probably plays a role during the productive stage of the HPV-16 life cycle [44]. The E5 protein may contribute to cellular transformation and prevent cell

death at the early stages of viral infection [45, 46]. It may also increase the carcinogenic activity of E7 [47, 48].

Oestrogen also activates cyclin-dependent kinases, supporting cell growth. Indole derivatives on the other hand, inhibit cyclin-dependent kinases and change the ratio of oestrogen metabolites in favour of 2-hydroxy-oestradiol, with resultant inhibition of cycle progression (see Fig. 2).

At a cellular level oestrogen appears to interact with HPV although little is known about the mechanisms at this level. Oestrogen inhibits apoptosis in cervical cancer cells [49, 50]. HPV oncogene expression may be increased in response to oestrogen and progesterone stimulation, thus causing cell growth [49, 51–57]. However, a study on HPV-16 positive CaSki and SiHa cells showed increased cell proliferation under the influence of oestrogen as well as progesterone but no change in HPV-16 E6/E7 transcription levels [50]. The authors suggested that the anti-apoptotic effect of oestrogen might allow a growth advantage for HPV infected cells [50]. In a model using luciferase reporter constructs under control of oestrogen response element promoter it was shown that HPV-16 E7 can dysregulate and reduce the function of steroid receptor coactivator 1, which may explain the role of steroid hormones as co-factors in the carcinogenesis of HR-HPV related disease [58]. In specimens from HPV positive cervical carcinomas increased levels of aromatase, an enzyme which converts androgens to oestrogen, were identified. This was associated with increased oestrogen receptor levels as well as

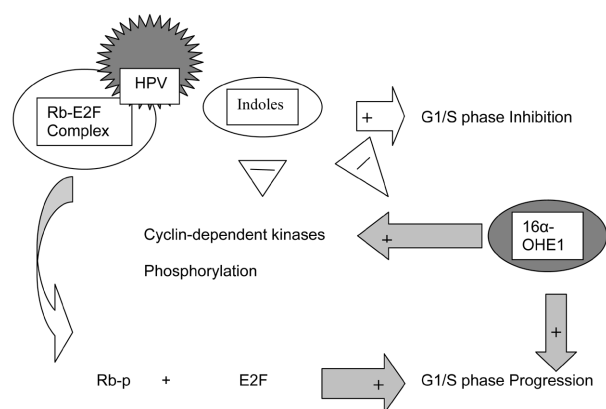


Figure 2. Schematic Illustration of the Effects of Oestrogen in HPV Infected Cells on the Tumour Suppressor Rb and Chemopreventive Action of Indole Derivates. HPV E7 oncoprotein binds to the tumour suppressor protein pRb and releases the E2F-transcription factor. Degraded pRb and free E2F can mediate the cell progress into the S phase. The released transcription factor E2F activates factors which promote cell-cycle progression. The oestrogen, 16 α -hydroxy-oestrone, also activates the cyclin-dependent kinases (CDKs) and thus supports cell progression into the S phase. Indole derivatives on the other hand reduce the 16 α -hydroxy-oestrone level and inhibit the cyclin-dependent kinases. Thus, Rb remains in an active state and cell progression is inhibited. Activation is symbolised by \Rightarrow , inhibition is symbolised by \triangle . Gray: potential malignant conversion, white: potential anti-proliferative effect.

expression of the HPV oncogenes E6 and E7 [59]. However, in specimens from women with cervical intraepithelial neoplasia a down-regulation of oestrogen-receptor has been observed [34, 60]. Other researchers suggest that oestrogen can increase the level of apoptosis associated with HPV-16 E2 protein [38]. Studies using HPV-transgenic mice suggested a permissive effect of oestrogen on cervical carcinogenesis as well as on tumour persistence and growth [27, 61–64]. A case-control study in 390 postmenopausal women suggested an increased risk of any HPV infection in past-users of combined hormone replacement therapy (HRT) depending on the duration of treatment [65]. However, the incidence of HR-HPV infection was low at 2.8% and there was no association between HPV infection and duration of treatment in current HRT users. Larger HRT studies did not demonstrate an increase in cervical cancer in HRT users [66, 67]. A meta-analysis suggested that the oral contraceptive pill could be a co-factor in HPV-related cervical carcinogenesis with an OR for the use of oral contraception and increased risk of cervical cancer of 2.82 (95%CI, 1.46–5.42) for five to nine years of use and 4.03 (95%CI, 2.09–8.02) for ten years of use and longer [16]. However, the ORs are low compared to the ORs of HR-HPV infection.

3 Chemoprevention with indoles

A variety of potential anticarcinogenic and chemopreventive agents have been examined over the last 30 years [68, 69]. Several epidemiological studies have demonstrated health benefits and cancer prevention for a “green” diet (for review see [69–71]). The evidence from epidemiological studies show conflicting results for an association between indole-derivates and cervical cancer [69]. Indoles, found in Brassicas (cruciferous vegetables such as cabbage, cauliflower, Brussels sprouts, kali, broccoli) have been shown to have chemopreventive properties. Indole-3-Carbinol (I3C) forms condensation products during acid digestion, of which diindolylmethane (DIM) appears to be one of the most active [72, 73].

Various antitumour and chemopreventive properties for indoles have been described *in vivo* and *in vitro* [69] including induction of apoptosis [74–76], cytotoxic effects [77, 78], cell cycle arrest [79–81], antioxidant effects [82–85], antiproliferative characteristics [82, 86], modulation of cell cycle parameters, activation of tumour suppressor genes such as BRCA1 and BRCA2 [87] and various cell signaling pathways, as well as causing a change in androgen and oestrogen metabolism. Micro-array analysis has demonstrated that I3C and DIM change the function of around 700 genes [88], some of which apparently reduce the expression of HPV oncogenes [85]. As HPV cannot be grown *in vitro*, experiments have been performed with cancer cell lines containing integrated HPV DNA rather than episomal DNA. Therefore extrapolation of the results has to be undertaken with caution when considering the effects of indole derivatives on HPV in an organic system.

4 Indoles and oestrogen metabolism

Degradation of 17 β -oestradiol, results in either 2-hydroxy-oestradiol, 16 α -hydroxy-oestrone or 4-hydroxy-oestrone metabolites (see Fig. 3). 2-hydroxy-oestradiol is anti-oestrogenic and anti-proliferative, whereas 16 α -hydroxy-oestrone and 4-hydroxy-oestrone cause cell proliferation and have carcinogenic potential (see reviews in [89, 90]).

Indole derivatives appear to influence oestrogen metabolism and control the growth of oestrogen-dependent cancer cells [54, 91–98], blocking abnormal proliferation and malignant transformation caused by oestradiol and 16 α -hydroxy-oestrone [99, 100]. Additionally, Indoles modulate oestrogen-receptor activities [87, 94, 101–104] as noted in breast cancer cell lines [94, 105], HPV-immortalised cell lines, cervical cancer cell lines [75, 98] and in a mouse model [61]. Mice expressing transgenes for HPV-16 develop cervical cancer when given 0.165 mg daily of 17-beta-estradiol over two months, whereas co-administration of 2000 ppm I3C results in a significant reduction in cervical cancers [61, 75]. The oestrogenic effects in this model

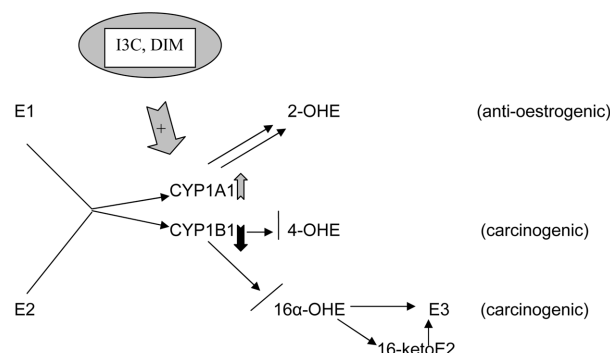


Figure 3. Schematic Simplified Illustration of Effect of Indole Derivates on Oestrogen Metabolism. Degradation of 17 β -oestradiol (E2) and oestrone (E1) by CYP1A1 results in 2-hydroxy-oestradiol (2-OHE) which has anti-oestrogenic and anti-proliferative properties. Degradation by CYP1B1 results in 16 α -hydroxy-oestrone (16-OHE) or 4-hydroxy-oestrone (4-OHE). Both cause cell proliferation and have carcinogenic potential. Indole derivates activate CYP1A1 but inhibit CYP1B1. This changes the balance in favour of 2-OHE production. At the same time, I3C can down-regulate the oestrogen-receptor α . Increase in metabolizing enzymes CYP1A1 is symbolized by \uparrow , decrease in metabolizing enzymes CYP1B1 is symbolized by \downarrow . E1, estrone; E2, 17 β -oestradiol; E3 oestrone; 4-OHE, 4-hydroxy-estrone; 16 α -OHE, 16 α -hydroxyestrone.

have to be interpreted with some caution as a keratin 14 promoter directed the HPV-16 gene expression rather than the native HPV promoter (K14-HPV 16 mice).

It is thought that the effects of Indoles on oestrogen metabolism are partially due to induction of enzymes (CYP1A1 family), critical for the metabolic detoxification of carcinogens [94, 106–111]. Activation of these liver enzymes increases hydroxylation of 17 β -oestradiol, leading to the protective 2-hydroxy oestradiol [54, 112–114].

In humans, excretion of 2-hydroxy oestradiol relative to that of 16 α -hydroxy oestrone was significantly increased by I3C [90, 100, 115–121]. Indole derivates also decrease CYP1B1 activity, thereby decreasing the formation of the carcinogenic 4-hydroxy oestrone (see review [89]). Not all humans respond to Indole derivates possibly explained by genetic polymorphic variation in CYP1A1 [122, 123].

5 Indoles and HPV

Indoles appear to have antiviral activity, reducing endogenous transcription of HPV16 oncogenes *in vitro* [85] and in K14-HPV16 mice [61].

A prospective trial utilising 400 mg I3C daily as treatment for recurrent respiratory papillomatosis in 33 patients showed that one third of patients experienced remission of papillomatous growth and did not require surgery whilst on I3C; one third had a reduction in papillomatous growth that

resulted in less frequent surgery and one third had no clinical response. No long-term side effects were observed [124]. Indole derivates are currently used as adjuvant treatment for recurrent respiratory papillomatosis.

A randomised placebo-controlled study of thirty patients with biopsy proven CIN II–III showed a 50% regression rate in women in the I3C arm (200 mg or 400 mg daily for three months), compared to none of the patients in the placebo arm (95% CI, 0.25 to 0.99; $p = 0.023$) [90].

6 Critics of indoles

Some researchers have noted oestrogenic properties for DIM [102, 105, 125], with DIM exhibiting both oestrogenic as well as anti-proliferative and cytotoxic activity [91, 102]. In rodent models I3C but not DIM appeared to enhance the hepatotoxicity of tamoxifen [114, 126]. Long-term intake of indole derivates may cause bone loss (unpublished data [127]).

Under certain conditions in cell and in animal studies indoles have been found to promote or enhance effects on tumourgenesis [128–136]. Some of the most critical voices come from studies with rainbow trout where indole derivates act as oestrogen agonists [125, 136] as in rat models [126, 137]. High indole intake appears to reduce immune responses by reducing natural killer cell activity [138, 139].

7 Safety and side effects

Recent animal safety studies have demonstrated increased safety for formulated DIM compared to I3C with no evidence of teratogenicity [140–144]. The doses given in animal studies are significantly higher than the relative doses given in human research [144]. Known side effects in human subjects are mild, reversible and uncommon and include nausea, dyspepsia and flatulence. Harmless darkening of the urine due to excretion of coloured DIM metabolites in those with low fluid intake is reported. There is also a suggestion that DIM may increase the severity of headaches in women who suffer migraine [145]. Long-term studies of the effects of indoles in humans are outstanding.

8 Conclusion

Many studies have investigated the potential anti-carcinogenic properties of indoles including their effects upon apoptosis, oestrogen and androgen metabolism. I3C modulates the oestrogen-receptor and enhances the activity of detoxifying enzymes responsible for 2-hydroxylation of oestrogen, inducing the formation of 2-hydroxy oestradiol, which in itself has anti-oestrogenic activities. The accumulated evidence supports a role for indoles as chemopreven-

tive agents in oestrogen-dependent tissue. Further long-term studies of chemoprevention for hormone-dependent tumours such as breast, cervical and prostate cancer in humans are warranted.

Both authors are involved in a chemoprevention study using DIM in women with low grade cervical cytological abnormalities. The study is supported by Cancer Research UK and BioResponse DIM®. Neither organisation has had an input into the present review.

Conflict of interest statement: Both authors are involved in a chemoprevention study using Diindolylmethane in women with low grade cervical cytological abnormalities. The study is supported by Cancer Research UK and BioResponse DIM®. Neither organisation has had an input into the present review.

9 References

- [1] zur Hausen, H., Papillomaviruses and cancer: From basic studies to clinical application, *Nat. Rev. Cancer* 2002, 2, 342–350.
- [2] International Agency for Research of Cancer Human Papillomavirus, *IARC Monographs on the Evaluation of Carcinogenic Risks to Humans*, Volume 64, Lyon 1995.
- [3] Munoz, N., Bosch, F. X., de Sanjose, S., Herrero, R. *et al.*, Epidemiologic classification of human papillomavirus types associated with cervical cancer. *N. Engl. J. Med.* 2003, 348, 518–527.
- [4] Bohmer, G., van den Brule, A. J., Brummer, O., Meijer, C. L., Petry, K. U., No confirmed case of human papillomavirus DNA-negative cervical intraepithelial neoplasia grade 3 or invasive primary cancer of the uterine cervix among 511 patients, *Am. J. Obstet. Gynecol.* 2003, 189, 118–120.
- [5] Bosch, F. X., Lorincz, A., Munoz, N., Meijer, C. J., Shah, K. V., The causal relation between human papillomavirus and cervical cancer, *J. Clin. Pathol.* 2002, 55, 244–265.
- [6] Munoz, N., Bosch, F. X., Castellsague, X., Diaz, M. *et al.*, Against which human papillomavirus types shall we vaccinate and screen? The international perspective, *Int. J. Cancer* 2004, 111, 278–285.
- [7] Koyamatsu, Y., Yokoyama, M., Nakao, Y., Fukuda, K. *et al.*, A comparative analysis of human papillomavirus types 16 and 18 and expression of p53 gene and Ki-67 in cervical, vaginal, and vulvar carcinomas, *Gynecol. Oncol.* 2003, 90, 547–551.
- [8] Daling, J. R., Madeleine, M. M., Johnson, L. G., Schwartz, S. M. *et al.*, Human papillomavirus, smoking, and sexual practices in the etiology of anal cancer, *Cancer* 2004, 101, 270–280.
- [9] Walboomers, J. M., Jacobs, M. V., Manos, M. M., Bosch, F. X. *et al.*, Human papillomavirus is a necessary cause of invasive cervical cancer worldwide, *J. Pathol.* 1999, 189, 12–19.
- [10] Kreimer, A. R., Clifford, G. M., Boyle, P., Franceschi, S., Human papillomavirus types in head and neck squamous cell carcinomas worldwide: a systematic review, *Cancer Epidemiol. Biomarkers Prev.* 2005, 14, 467–475.
- [11] Syrjanen, S., Human papillomavirus (HPV) in head and neck cancer, *J. Clin. Virol.* 2005, 32, S59–S66.
- [12] Major, T., Szarka, K., Sziklai, I., Gergely, L., Czegledy, J., The characteristics of human papillomavirus DNA in head and neck cancers and papillomas, *J. Clin. Pathol.* 2005, 58, 51–55.
- [13] Gerein, V., Rastorguev, E., Gerein, J., Draf, W., Schirren, J., Incidence, age at onset, and potential reasons of malignant transformation in recurrent respiratory papillomatosis patients: 20 years experience, *Otolaryngol. Head Neck Surg.* 2005, 132, 392–394.
- [14] Plummer, M., Herrero, R., Franceschi, S., Meijer, C. J. *et al.*, Smoking and cervical cancer: pooled analysis of the IARC multi-centric case–control study, *Cancer Causes Control* 2003, 14, 805–814.
- [15] Vessey, M., Painter, R., Yeates, D., Mortality in relation to oral contraceptive use and cigarette smoking, *Lancet* 2003, 362, 185–191.
- [16] Moreno, V., Bosch, F. X., Munoz, N., Meijer, C. J. *et al.*, Effect on oral contraceptives in women with human papillomavirus infection: For the International Agency for Research on Cancer (IARC). Multicentric Cervical Study Group, *Lancet* 2002, 359, 1085–1092.
- [17] Hinkula, M., Pukkala, E., Kyyronen, P., Laukkanen, P. *et al.*, A population-based study on the risk of cervical cancer and cervical intraepithelial neoplasia among grand multiparous women in Finland, *Br. J. Cancer* 2004, 90, 1025–1029.
- [18] Hildesheim, A., Herrero, R., Castle, P. E., Wacholder, S. *et al.*, HPV co-factors related to the development of cervical cancer: Results from a population-based study in Costa Rica, *Br. J. Cancer* 2001, 84, 1219–1226.
- [19] Smith, J. S., Bosetti, C., Munoz, N., Herrero, R. *et al.*, Chlamydia trachomatis and invasive cervical cancer: a pooled analysis of the International Agency for Research on Cancer multicentric case-control study, *Int. J. Cancer* 2004, 111, 431–439.
- [20] Matsumoto, K., Yasugi, T., Oki, A., Hoshiai, H. *et al.*, Are smoking and chlamydial infection risk factors for CIN? Different results after adjustment for HPV DNA and antibodies, *Br. J. Cancer* 2003, 89, 831–833.
- [21] Koskela, P., Anttila, T., Bjorge, T., Brunsvig, A. *et al.*, Chlamydia trachomatis infection as a risk factor for invasive cervical cancer, *Int. J. Cancer* 2000, 85, 35–39.
- [22] Hakama, M., Luostarinen, T., Hallmans, G., Jellum, E. *et al.*, Joint effect of HPV16 with Chlamydia trachomatis and smoking on risk of cervical cancer: Antagonism or misclassification (Nordic countries), *Cancer Causes Control* 2000, 11, 783–790.
- [23] Peh, W. L., Middleton, K., Christensen, N., Nicholls, P. *et al.*, Life cycle heterogeneity in animal models of human papillomavirus-associated disease, *J. Virol.* 2002, 76, 10401–10416.
- [24] Singer, A., Monaghan, J., Quek, S. C., Deery, A., in: Singer, A. Monaghan, J., (Eds.), *Lower Genital Tract Precancer: Colposcopy, Pathology and Treatment*, Blackwell Science, London 2000, pp. 1–12, 71–95.
- [25] Autier, P., Coibion, M., Huet, F., Grivegne, A. R., Transformation zone location and intraepithelial neoplasia of the cervix uteri, *Br. J. Cancer* 1996, 74, 488–490.
- [26] Elson, D. A., Riley, R. R., Lacey, A., Thordarson, G. *et al.*, Sensitivity of the cervical transformation zone to estrogen-induced squamous carcinogenesis, *Cancer Res.* 2000, 60, 1267–1275.

- [27] Brake, T., Lambert, P. F., Estrogen contributes to the onset, persistence, and malignant progression of cervical cancer in a human papillomavirus-transgenic mouse model, *Proc. Natl. Acad. Sci. USA* 2005, 102, 2490–2495.
- [28] Remoue, F., Jacobs, N., Miot, V., Boniver, J., Delvenne, P., High intraepithelial expression of estrogen and progesterone receptors in the transformation zone of the uterine cervix, *Am. J. Obstet. Gynecol.* 2003, 189, 1660–1665.
- [29] Monsonogo, J., Magdelenat, H., Catalan, F., Coscas, Y. *et al.*, Estrogen and progesterone receptors in cervical human papillomavirus related lesions, *Int. J. Cancer* 1991, 48, 533–539.
- [30] Gonzalez Sanchez, J. L., Chavez Brambila, J., Maricela Roman, A., Infante Martinez, R., Salazar Esquivel, L. E., Value of estrogen and progesterone receptors in the management of intraepithelial squamous lesions of low grade, *Ginecol. Obstet. Mex.* 2001, 69, 1–5.
- [31] Shew, M. L., McGlennen, R., Zaidi, N., Westerheim, M. *et al.*, Oestrogen receptor transcripts associated with cervical human papillomavirus infection, *Sex. Transm. Infect.* 2002, 78, 210–214.
- [32] Fonseca-Moutinho, J. A., Cruz, E., Carvalho, L., Prazeres, H. J. *et al.*, Estrogen receptor, progesterone receptor, and bcl-2 are markers with prognostic significance in CIN III, *Int. J. Gynecol. Cancer* 2004, 14, 911–920.
- [33] Coelho, F. R., Prado, J. C., Pereira Sobrinho, J. S., Hamada, G. *et al.*, Estrogen and progesterone receptors in human papilloma virus-related cervical neoplasia, *Braz. J. Med. Biol. Res.* 2004, 37, 83–88.
- [34] Tervahauta, A., Syrjanen, S., Syrjanen, K., Epidermal growth factor receptor, c-erbB-2 proto-oncogene and estrogen receptor expression in human papillomavirus lesions of the uterine cervix, *Int. J. Gynecol. Pathol.* 1994, 13, 234–240.
- [35] Shen, K., Yueng, W., Ngan, H., Estrogen and progesterone receptors in normal cervix and primary cervical carcinoma, *Chin. Med. J. (Engl)* 1994, 107, 648–652.
- [36] Nonogaki, H., Fujii, S., Konishi, I., Nanbu, Y. *et al.*, Estrogen receptor localization in normal and neoplastic epithelium of the uterine cervix, *Cancer* 1990, 66, 2620–2627.
- [37] Konishi, I., Fujii, S., Nonogaki, H., Nanbu, Y. *et al.*, Immunohistochemical analysis of estrogen receptors, progesterone receptors, Ki-67 antigen, and human papillomavirus DNA in normal and neoplastic epithelium of the uterine cervix, *Cancer* 1991, 68, 1340–1350.
- [38] Webster, K., Taylor, A., Gaston, K., Oestrogen and progesterone increase the levels of apoptosis induced by the human papillomavirus type 16 E2 and E7 proteins, *J. Gen. Virol.* 2001, 82, 201–213.
- [39] Doorbar, J., Molecular biology of human papillomavirus infection and cervical cancer, *Clin. Sci. (Lond.)* 2006, 110, 525–541.
- [40] Hengstermann, A., D'Silva, M. A., Kuballa, P., Butz, K. *et al.*, Growth suppression induced by downregulation of E6-AP expression in human papillomavirus-positive cancer cell lines depends on p53, *J. Virol.* 2005, 79, 9296–9300.
- [41] Thomas, M., Banks, L., Inhibition of Bak-induced apoptosis by HPV-18 E6, *Oncogene* 1998, 17, 2943–2954.
- [42] Vogt, M., Butz, K., Dymalla, S., Semzow, J., Hoppe-Seyler, F., Inhibition of Bax activity is crucial for the antiapoptotic function of the human papillomavirus E6 oncoprotein, *Oncogene* 2006, 25, 4009–4015.
- [43] Magal, S. S., Jackman, A., Ish-Shalom, S., Botzer, L. E. *et al.*, Downregulation of Bax mRNA expression and protein stability by the E6 protein of human papillomavirus 16, *J. Gen. Virol.* 2005, 86, 611–621.
- [44] Genther, S. M., Sterling, S., Duensing, S., Munger, K. *et al.*, Quantitative role of the human papillomavirus type 16 E5 gene during the productive stage of the viral life cycle, *J. Virol.* 2003, 77, 2832–2842.
- [45] Kabsch, K., Mossadegh, N., Kohl, A., Komposch, G. *et al.*, The HPV-16 E5 protein inhibits TRAIL- and FasL-mediated apoptosis in human keratinocyte raft cultures, *Intervirology* 2004, 47, 48–56.
- [46] Auvinen, E., Alonso, A., Auvinen, P., Human papillomavirus type 16 E5 protein colocalizes with the antiapoptotic Bcl-2 protein, *Arch. Virol.* 2004, 149, 1745–1759.
- [47] Bouvard, V., Matlaszewski, G., Gu, Z. M., Storey, A., Banks, L., The human papillomavirus type 16 E5 gene cooperates with the E7 gene to stimulate proliferation of primary cells and increases viral gene expression, *Virology* 1994, 203, 73–80.
- [48] Crusius, K., Rodriguez, I., Alonso, A., The human papillomavirus type 16 E5 protein modulates ERK1/2 and p38 MAP kinase activation by an EGFR-independent process in stressed human keratinocytes, *Virus Gene* 2000, 20, 65–99.
- [49] Chen, D., Carter, T. H., Auborn, K. J., Apoptosis in cervical cancer cells: implications for adjunct anti-estrogen therapy for cervical cancer, *Anticancer Res.* 2004, 24, 2649–2656.
- [50] Ruutu, M., Wahlroos, N., Syrjanen, K., Johansson, B., Syrjanen, S., Effects of 17beta-estradiol and progesterone on transcription of human papillomavirus 16 E6/E7 oncogenes in CaSki and SiHa cell lines, *Int. J. Gynecol. Cancer* 2006, 16, 1261–1268.
- [51] Kim, C. J., Um, T. Y., Kim, E. J., Park, T. C. *et al.*, Regulation of cell growth and HPV genes by exogenous estrogen in cervical cancer cells, *Int. J. Gynecol. Cancer* 2000, 10, 157–164.
- [52] Mitrani-Rosenbaum, S., Tsvieli, R., Tur-Kaspa, R., Oestrogen stimulates differential transcription of human papillomavirus type 16 in SiHa cervical carcinoma cells, *J. Gen. Virol.* 1989, 70, 2227–2232.
- [53] Auborn, K. J., Woodworth, C., DiPaolo, J. A., Bradlow, H. L., The interaction between HPV infection and estrogen metabolism in cervical carcinogenesis, *Int. J. Cancer* 1991, 49, 867–869.
- [54] Yuan, F., Chen, D. Z., Liu, K., Sepkovic, D. W. *et al.*, Anti-estrogenic activities of indole-3-carbinol in cervical cells: implication for prevention of cervical cancer, *Anticancer Res.* 1999, 19, 1673–1680.
- [55] Michelin, D., Gissmann, L., Street, D., Potkul, R. K. *et al.*, Regulation of human papillomavirus type 18 *in vivo*, effects of estrogen and progesterone in transgenic mice, *Gynecol. Oncol.* 1997, 66, 20–28.
- [56] Yuan, F., Auborn, K. J., James, C., Altered growth and viral gene expression in human papillomavirus type 16-containing cancer cell lines treated with progesterone, *Cancer Invest.* 1999, 17, 19–29.
- [57] Kim, C. J., Um, S. J., Kim, T. Y., Kim, E. J. *et al.*, Regulation of cell growth and HPV genes by exogenous estrogen in cervical cancer cells, *Int. J. Gynecol. Cancer* 2000, 10, 157–164.
- [58] Baldwin, A., Huh, K. W., Munger, K., Human papillomavirus E7 oncoprotein dysregulates steroid receptor coactivator 1 localization and function, *J. Virol.* 2006, 80, 6669–6677.

- [59] Nair, H. B., Luthra, R., Kirma, N., Liu, Y. G. *et al.*, Induction of aromatase expression in cervical carcinomas: effects of endogenous estrogen on cervical cancer cell proliferation, *Cancer Res.* 2005, 65, 11164–11173.
- [60] Bekkers, R. L., van der Avoort, I. A., Melchers, W. J., Bulten, J. *et al.*, Down regulation of estrogen receptor expression is an early event in human papillomavirus infected cervical dysplasia, *Eur. J. Gynaecol. Oncol.* 2005, 26, 376–382.
- [61] Jin, L., Qi, M., Chen, D. Z., Anderson, A. *et al.*, Indole-3-carbinol prevents cervical cancer in human papilloma virus type 16 (HPV16) transgenic mice, *Cancer Res.* 1999, 59, 3991–3997.
- [62] Riley, R. R., Duensing, S., Brake, T., Munger, K. *et al.*, Dissection of human papillomavirus E6 and E7 function in transgenic mouse models of cervical carcinogenesis, *Cancer Res.* 2003, 63, 4862–4871.
- [63] Arbeit, J. M., Howley, P. M., Hanahan, D., Chronic estrogen-induced cervical and vaginal squamous carcinogenesis in human papillomavirus type 16 transgenic mice, *Proc. Natl. Acad. Sci. USA* 1996, 93, 2930–2935.
- [64] Shai, A., Brake, T., Somoza, C., Lambert, P. F., The human papillomavirus E6 oncogene dysregulates the cell cycle and contributes to cervical carcinogenesis through two independent activities, *Cancer Res.* 2007, 67, 1626–1635.
- [65] Kutza, J., Smith, E., Levy, B., Jian, J. *et al.*, Use of hormone replacement therapy (HRT) and detection of human papillomavirus (HPV) DNA in postmenopausal women, *Ann. Epidemiol.* 2000, 10, 465–466.
- [66] Beral, V., Banks, E., Reeves, G., Evidence from randomised trials on the long-term effects of hormone replacement therapy, *Lancet* 2002, 360, 942–944.
- [67] Rossouw, J. E., Anderson, G. L., Prentice, R. L., LaCroix, A. Z. *et al.*, Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial, *JAMA* 2002, 288, 321–333.
- [68] Chakraborty, S., Roy, M., Bhattacharya, R. K., Prevention and repair of DNA damage by selected phytochemicals as measured by single cell gel electrophoresis, *J. Environ. Pathol. Toxicol. Oncol.* 2004, 23, 215–226.
- [69] International Agency for Research of Cancer. Cruciferous vegetables, Isothiocyanates and Indoles (Handbook of Cancer Prevention, Volume 9), IARC Press, Lyon 2004.
- [70] Temple, N. J., Gladwin, K. K., Fruit, vegetables, and the prevention of cancer: research challenges, *Nutrition* 2003, 19, 467–470.
- [71] Verhoeven, D. T., Verhagen, R. A., Goldbohm, R. A., van den Brandt, P. A., van Poppel, G., A review of mechanisms underlying anticarcinogenicity by brassica vegetables, *Chem. Biol. Interact.* 1997, 103, 79–129.
- [72] Arneson, D. W., Hurwitz, A., McMahon, L. M., Robaugh, D., Presence of 3'-Diindolylmethane in human plasma after oral administration of indole-3-carbinol, *Proc. Am. Assoc. Cancer Res.* 1999, 40, 429.
- [73] Sepkovic, D. W., Bradlow, H. L., Bell, M., Quantitative determination of 3,3'-diindolylmethane in urine of individuals receiving indole-3-carbinol, *Nutr. Cancer* 2001, 41, 57–63.
- [74] Nachshon-Kedmi, M., Yannai, S., Fares, F. A., Induction of apoptosis in human prostate cancer cell line, PC3, by 3,3'-diindolylmethane through the mitochondrial pathway, *Br. J. Cancer* 2004, 91, 1358–1363.
- [75] Chen, D. Z., Qi, M., Auborn, K. J., Carter, T. H., Indole-3-carbinol and diindolylmethane induce apoptosis of human cervical cancer cells and in murine HPV16-transgenic preneoplastic cervical epithelium, *J. Nutr.* 2001, 131, 3294–3302.
- [76] Abdelrahim, M., Newman, K., Vanderlaag, K., Samudio, I., Safe, S., 3,3'-Diindolylmethane (DIM) and its derivatives induce apoptosis in pancreatic cancer cells through endoplasmic reticulum stress-dependent upregulation of DR5, *Carcinogenesis* 2006, 27, 717–728.
- [77] Sun, S., Han, J., Ralph, W. M., Jr., Chandrasekaran, A. *et al.*, Endoplasmic reticulum stress as a correlate of cytotoxicity in human tumor cells exposed to diindolylmethane *in vitro*, *Cell Stress Chaperones* 2004, 9, 76–87.
- [78] Pappa, G., Lichtenberg, M., Iori, R., Barillari, J. *et al.*, Comparison of growth inhibition profiles and mechanisms of apoptosis induction in human colon cancer cell lines by isothiocyanates and indoles from *Brassicaceae*, *Mutat. Res.* 2006, 599, 76–87.
- [79] Hong, C., Kim, H. A., Firestone, G. L., Bjeldanes, L. F., 3,3'-Diindolylmethane induces a G1 cell cycle arrest in human breast cancer cells that is accompanied by Sp1-mediated activation of p21 expression, *Carcinogenesis* 2002, 23, 1297–1305.
- [80] Firestone, G. L., Bjeldanes, L. F., Indole-3-carbinol and 3,3'-diindolylmethane antiproliferative signaling pathways control cell-cycle gene transcription in human breast cancer cells by regulating promoter-Sp1 transcription factor interactions, *J. Nutr.* 2003, 133, 2448S–2455S.
- [81] Gong, Y., Firestone, G. L., Bjeldanes, L. F., 3,3'-Diindolylmethane Is a novel topoisomerase II (alpha) catalytic inhibitor that induces S-phase retardation and mitotic delay in human hepatoma HepG2 cells, *Mol. Pharmacol.* 2006, 69, 1320–1327.
- [82] Benabadi, S. H., Wen, R., Zheng, J. B., Dong, X. C., Yuan, S. G., Anticarcinogenic and antioxidant activity of diindolylmethane derivatives, *Acta Pharmacol. Sin.* 2004, 25, 666–671.
- [83] Arnao, M. B., Sanchez-Bravo, J., Acosta, M., Indole-3-carbinol as a scavenger of free radicals, *Biochem. Mol. Biol. Int.* 1996, 39, 1125–1135.
- [84] Shertzer, H. G., Berger, M. L., Tabor, M. W., Intervention in free radical mediated hepatotoxicity and lipid peroxidation by indole-3-carbinol, *Biochem. Pharmacol.* 1988, 37, 333–338.
- [85] Carter, T. H., Liu, K., Ralph, W., Jr., Chen, D. *et al.*, Diindolylmethane alters gene expression in human keratinocytes *in vitro*, *J. Nutr.* 2002, 132, 3314–3324.
- [86] Garikapaty, V. P., Ashok, B. T., Tadi, K., Mittelman, A., Tiwari, R. K., 3,3'-Diindolylmethane downregulates pro-survival pathway in hormone independent prostate cancer, *Biochem. Biophys. Res. Commun.* 2006, 340, 718–725.
- [87] Fan, S., Meng, Q., Auborn, K., Carter, T., Rosen, E. M., BRCA1 and BRCA2 as molecular targets for phytochemicals indole-3-carbinol and genistein in breast and prostate cancer cells, *Br. J. Cancer* 2006, 94, 407–426.
- [88] Li, Y., Li, X., Sarkar, F. H., Gene expression profiles of I3C- and DIM-treated PC3 human prostate cancer cells determined by cDNA microarray analysis, *J. Nutr.* 2003, 133, 1011–1019.
- [89] Bradlow, H. L., Sepkovic, D. W., Telang, N. T., Osborne, M. P., Multifunctional aspects of the action of indole-3-carbinol as an antitumor agent, *Ann. NY Acad. Sci.* 1999, 889, 204–213.

- [90] Bell, M. C., Crowley-Nowick, P., Bradlow, H. L., Sepkovic, D. W. *et al.*, Placebo-controlled trial of indole-3-carbinol in the treatment of CIN, *Gynecol. Oncol.* 2000, 78, 123–129.
- [91] Leong, H., Firestone, G. L., Bjeldanes, L. F., Cytostatic effects of 3,3'-diindolylmethane in human endometrial cancer cells result from an estrogen receptor-mediated increase in transforming growth factor- α expression, *Carcinogenesis* 2001, 22, 1809–1817.
- [92] Chen, I., McDougal, A., Wang, F., Safe, S., Aryl hydrocarbon receptor-mediated antiestrogenic and antitumorigenic activity of diindolylmethane, *Carcinogenesis* 1998, 19, 1631–1639.
- [93] Tiwari, R. K., Guo, L., Bradlow, H. L., Telang, N. T., Osborne, M. P., Selective responsiveness of human breast cancer cells to indole-3-carbinol, a chemopreventive agent, *J. Natl. Cancer Inst.* 1994, 86, 126–131.
- [94] Ashok, B. T., Chen, Y., Liu, X., Bradlow, H. L. *et al.*, Abrogation of estrogen-mediated cellular and biochemical effects by indole-3-carbinol, *Nutr. Cancer* 2001, 41, 180–187.
- [95] Kociba, R. J., Keyes, D. G., Beyer, J. E., Carreon, R. M. *et al.*, Results of a two-year chronic toxicity and oncogenicity study of 2,3,7,8-tetrachlorodibenzo-p-dioxin in rats, *Toxicol. Appl. Pharmacol.* 1978, 46, 279–303.
- [96] Wattenberg, L. W., Loub, W. D., Inhibition of polycyclic aromatic hydrocarbon-induced neoplasia by naturally occurring indoles, *Cancer Res.* 1978, 38, 1410–1413.
- [97] Grubbs, C. J., Steele, V. E., Casebolt, T., Juliana, M. M. *et al.*, Chemoprevention of chemically-induced mammary carcinogenesis by indole-3-carbinol, *Anticancer Res.* 1995, 15, 709–716.
- [98] Newfield, L., Bradlow, H. L., Sepkovic, D. W., Auburn, K., Estrogen metabolism and the malignant potential of human papillomavirus immortalized keratinocytes, *Proc. Soc. Exp. Biol. Med.* 1998, 217, 322–326.
- [99] Newfield, L. B. H., Sepkovic, D. W., Auburn, K., Estrogen metabolism and the malignant potential of human papillomavirus immortalized keratinocytes, *Proc. Soc. Exp. Biol. Med.* 1998, 217, 322–326.
- [100] Telang, N. T., Katdare, M., Bradlow, H. L., Osborne, M. P., Fishman, J., Inhibition of proliferation and modulation of estradiol metabolism: novel mechanisms for breast cancer prevention by the phytochemical indole-3-carbinol, *Proc. Soc. Exp. Biol. Med.* 1997, 216, 246–252.
- [101] Meng, Q., Yuan, F., Goldberg, I. D., Rosen, E. M. *et al.*, Indole-3-carbinol is a negative regulator of estrogen receptor- α signaling in human tumor cells, *J. Nutr.* 2000, 130, 2927–2931.
- [102] Leong, H., Riby, J. E., Firestone, G. L., Bjeldanes, L. F., Potent ligand-independent estrogen receptor activation by 3,3'-diindolylmethane is mediated by cross talk between the protein kinase A and mitogen-activated protein kinase signaling pathways, *Mol. Endocrinol.* 2004, 18, 291–302.
- [103] Wang, T. T., Milner, M. J., Milner, J. A., Kim, Y. S., Estrogen receptor α as a target for indole-3-carbinol, *J. Nutr. Biochem.* 2006, 10, 659–664.
- [104] Auburn, K. J., Fan, S., Rosen, E. M., Goodwin, L. *et al.*, Indole-3-carbinol is a negative regulator of estrogen, *J. Nutr.* 2003, 133, 2470S–2475S.
- [105] Riby, J. E., Chang, G. H., Firestone, G. L., Bjeldanes, L. F., Ligand-independent activation of estrogen receptor function by 3,3'-diindolylmethane in human breast cancer cells, *Biochem. Pharmacol.* 2000, 60, 167–177.
- [106] Chen, I., Safe, S., Bjeldanes, L., Indole-3-carbinol and diindolylmethane as aryl hydrocarbon (Ah) receptor agonists and antagonists in T47D human breast cancer cells, *Biochem. Pharmacol.* 1996, 51, 1069–1076.
- [107] Bradfield, C. A., Bjeldanes, L. F., High-performance liquid chromatographic analysis of anticarcinogenic indoles in brassica oleracea, *J. Agric. Food Chem.* 1987, 35, 46–49.
- [108] Bradfield, C. A., Bjeldanes, L. F., Structure-activity relationships of dietary indoles: a proposed mechanism of action as modifiers of xenobiotic metabolism, *J. Toxicol. Environ. Health* 1987, 21, 311–323.
- [109] Bjeldanes, L. F., Kim, J. Y., Grose, K. R., Bartholomew, J. C., Bradfield, C. A., Aromatic hydrocarbon responsiveness-receptor agonists generated from indole-3-carbinol in vitro and in vivo: comparisons with 2,3,7,8-tetrachlorodibenzo-p-dioxin, *Proc. Natl. Acad. Sci. USA* 1991, 88, 9543–9547.
- [110] Nho, C. W., Jeffery, E., The synergistic upregulation of phase II detoxification enzymes by glucosinolate breakdown products in cruciferous vegetables, *Toxicol. Appl. Pharmacol.* 2001, 174, 146–152.
- [111] Sanderson, J. T., Slobbe, L., Lansbergen, G. W., Safe, S., van den Berg, M., 2,3,7,8-Tetrachlorodibenzo-p-dioxin and diindolylmethanes differentially induce cytochrome P450 1A1, 1B1, and 19 in H295R human adrenocortical carcinoma cells, *Toxicol. Sci.* 2001, 61, 40–48.
- [112] Horn, T. L., Reichert, M. A., Bliss, R. L., Malejka-Giganti, D., Modulations of P450 mRNA in liver and mammary gland and P450 activities and metabolism of estrogen in liver by treatment of rats with indole-3-carbinol, *Biochem. Pharmacol.* 2002, 64, 393–404.
- [113] Ritter, C. L., Prigge, W. F., Reichert, M. A., Malejka-Giganti, D., Oxidations of 17 β -estradiol and estrone and their interconversions catalyzed by liver, mammary gland and mammary tumor after acute and chronic treatment of rats with indole-3-carbinol or beta-naphthoflavone, *Can. J. Physiol. Pharmacol.* 2001, 79, 519–532.
- [114] Parkin, D. R., Malejka-Giganti, D., Differences in the hepatic P450-dependent metabolism of estrogen and tamoxifen in response to treatment of rats with 3,3'-diindolylmethane and its parent compound indole-3-carbinol, *Cancer Detect. Prev.* 2004, 28, 72–79.
- [115] Wong, G. Y., Bradlow, L., Sepkovic, D., Mehl, S. *et al.*, Dose-ranging study of indole-3-carbinol for breast cancer prevention, *J. Cell Biochem.* 1997, 28–29, 111–116.
- [116] Dalessandri, K. M., Firestone, G. L., Fitch, M. D., Bradlow, H. L., Bjeldanes, L. F., Pilot study: effect of 3,3'-diindolylmethane supplements on urinary hormone metabolites in postmenopausal women with a history of early-stage breast cancer, *Nutr. Cancer* 2004, 50, 161–167.
- [117] Bradlow, H. L., Michnovicz, J. J., Halper, M., Miller, D. G. *et al.*, Long-term responses of women to indole-3-carbinol or a high fiber diet, *Cancer Epidemiol. Biomarkers Prev.* 1994, 3, 591–595.
- [118] Auburn, K., Abramson, A., Bradlow, H. L., Sepkovic, D., Mullooly, V., Estrogen metabolism and laryngeal papillomatosis: a pilot study on dietary prevention, *Anticancer Res.* 1998, 18, 4569–4573.
- [119] Bradlow, H. L., Davis, D. L., Lin, G., Sepkovic, D., Tiwari, R., Effects of pesticides on the ratio of 16 α /2-hydroxy-estrone: a biologic marker of breast cancer risk, *Environ. Health Perspect.* 1995, 103, 147–150.
- [120] Michnovicz, J. J., Bradlow, H. L., Altered estrogen metabolism and excretion in humans following consumption of indole-3-carbinol, *Nutr. Cancer* 1991, 16, 59–66.

- [121] Michnovicz, J. J., Adlercreutz, H., Bradlow, H. L., Changes in levels of urinary estrogen metabolites after oral indole-3-carbinol treatment in humans, *J. Natl. Cancer Inst.* 1997, **89**, 718–723.
- [122] Taioli, E., Trachman, J., Chen, X., Toniolo, P., Garte, S. J., A CYP1A1 restriction fragment length polymorphism is associated with breast cancer in African-American women, *Cancer Res.* 1995, **55**, 3757–3758.
- [123] Taioli, E., Bradlow, H. L., Garbers, S. V., Sepkovic, D. W. *et al.*, Role of estradiol metabolism and CYP1A1 polymorphisms in breast cancer risk, *Cancer Detect. Prev.* 1999, **23**, 232–237.
- [124] Rosen, C. A., Bryson, P. C., Indole-3-carbinol for recurrent respiratory papillomatosis: long-term results. *J. Voice* 2004, **18**, 248–253.
- [125] Shilling, A. D., Carlson, D. B., Katchamart, S., Williams, D. E., 3,3'-Diindolylmethane, a major condensation product of indole-3-carbinol, is a potent estrogen in the rainbow trout, *Toxicol. Appl. Pharmacol.* 2001, **170**, 191–200.
- [126] Crowell, J., A Indole-3-carbinol, but not its major digestive product 3,3'-diindolylmethane, induces reversible hepatocyte hypertrophy and cytochromes P450, *Toxicol. Appl. Pharmacol.* 2006, **211**, 115–123.
- [127] Auburn, K. J., Therapy for recurrent respiratory papillomatosis, *Antivir. Ther.* 2002, **7**, 1–9.
- [128] Bailey, G. S., Hendricks, J. D., Shelton, D. W., Nixon, J. E., Pawlowski, N. E., Enhancement of carcinogenesis by the natural anticarcinogen indole-3-carbinol, *J. Natl. Cancer Inst.* 1987, **78**, 931–934.
- [129] Dashwood, R. H., Fong, A. T., Williams, E. D., Hendricks, J. D., Bailey, G. S., Promotion of aflatoxin B1 carcinogenesis by the natural tumour modulator indole-3-carbinol: influence of dose, duration, and intermittent exposure on indole-3-carbinol promotional potency, *Cancer Res.* 1991, **51**, 2362–2365.
- [130] Kim, D. J., Lee, K. K., Han, B. S., Ahn, B. *et al.*, Biphasic modifying effect of indole-3-carbinol on diethylnitrosamine-induced preneoplastic glutathione S-transferase placental form-positive liver cell foci in Sprague-Dawley rats, *Jpn. J. Cancer Res.* 1994, **85**, 578–583.
- [131] Kim, D. J., Han, B. S., Ahn, B., Hasegawa, R. *et al.*, Enhancement by indole-3-carbinol of liver and thyroid gland neoplastic development in a rat medium-term multiorgan carcinogenesis model, *Carcinogenesis* 1997, **18**, 377–381.
- [132] Pence, B. C., Buddingh, F., Yang, S. P., Multiple dietary factors in the enhancement of dimethylhydrazine carcinogenesis: main effect of indole-3-carbinol, *J. Natl. Cancer Inst.* 1986, **77**, 269–276.
- [133] Dashwood, R. H., Xu, M., The disposition and metabolism of 2-amino-3-methylimidazo-[4,5-f]quinoline in the F344 rat at high versus low doses of indole-3-carbinol, *Food Chem. Toxicol.* 2003, **41**, 1185–1192.
- [134] Renwick, A. B., Mistry, H., Barton, P. T., Mallet, F. *et al.*, Effect of some indole derivatives on xenobiotic metabolism and xenobiotic-induced toxicity in cultured rat liver slices, *Food Chem. Toxicol.* 1999, **37**, 609–618.
- [135] Tilton, S. C., Givan, S. A., Pereira, C. B., Bailey, G. S., Williams, D. E., Toxicogenomic profiling of the hepatic tumor promoters indole-3-carbinol, 17 β -estradiol and beta-naphthoflavone in rainbow trout, *Toxicol. Sci.* 2006, **90**, 61–72.
- [136] Tilton, S. C., Hendricks, J. D., Orner, G. A., Pereira, C. B. *et al.*, Gene expression analysis during tumor enhancement by the dietary phytochemical, 3,3'-diindolylmethane, in rainbow trout, *Carcinogenesis* 2007, **28**, 1589–1598.
- [137] Yoshida, M., Katashima, S., Ando, J., Tanaka, T. *et al.*, Dietary indole-3-carbinol promotes endometrial adenocarcinoma development in rats initiated with N-ethyl-N'-nitro-N-nitrosoguanidine, with induction of cytochrome P450s in the liver and consequent modulation of estrogen metabolism, *Carcinogenesis* 2004, **25**, 2257–2264.
- [138] Exon, J. H., South, E. H., Magnuson, B. A., Hednrix, K., Effects of indole-3-carbinol on immune responses, aberrant crypt foci and colonic crypt cell proliferation in rats, *J. Toxicol. Environ. Health A* 2001, **62**, 561–573.
- [139] Exon, J. H., South, E. H., Dietary indole-3-carbinol alters immune functions in rats, *J. Toxicol. Environ. Health A* 2000, **59**, 271–279.
- [140] Johnson, W. D., McCormick, D. L., Crowell, J. A., A Range-Finding Developmental Study of Orally Administered Diindolylmethane in Rats. NCI Sponsored IITRI Study #1169, 2003.
- [141] Nishie, K., Daxenbichler, M. E., Toxicology of glucosinolates, related compounds (nitriles, R-goitrin, isothiocyanates) and vitamin U found in cruciferae, *Food Cosmet. Toxicol.* 1980, **18**, 159–172.
- [142] Gao, X., Petroff, B. K., Oluola, O., Georg, G. *et al.*, Endocrine disruption by indole-3-carbinol and tamoxifen: blockage of ovulation, *Toxicol. Appl. Pharmacol.* 2002, **183**, 179–188.
- [143] Crowell, J. A., Page, J. G., Levine, B. S., Tomlinson, M. J., Herbert, C. D., Indole-3-carbinol, but not its major digestive product 3,3'-diindolylmethane, induces reversible hepatocyte hypertrophy and cytochromes P450, *Toxicol. Appl. Pharmacol.* 2006, **211**, 115–123.
- [144] Leibelt, D. A., Hedstrom, O. R., Fischer, K. A., Pereira, C. B., Williams, D. E., Evaluation of chronic dietary exposure to indole-3-carbinol and absorption-enhanced 3,3'-diindolylmethane in sprague-dawley rats, *Toxicol. Sci.* 2003, **74**, 10–21.
- [145] Zeligs, M. A., Seplovic, D. W., Manrique, C. A., Macksalka, M. *et al.*, Absorption-enhanced 3,3'-diindolylmethane: Human use in HPV-related, benign and pre-cancerous conditions. *Proc. Am. Assoc. Cancer Res.* 2002, **43**, 644.